

REVIEW Open Access

Role of Wnt canonical pathway in hematological malignancies

Xueling GE, Xin Wang*

Abstract

Wnt canonical signaling pathway plays a diverse role in embryonic development and maintenance of organs and tissues in adults. It has been observed that $\text{Wnt/}\beta$ -catenin signaling pathway is involved in the pathogenesis of many carcinomas. Moreover, $\text{Wnt/}\beta$ -catenin pathway has been revealed to be associated with angiogenesis. Wnt canonical pathway signaling has great potential as a therapeutic target. It has been disclosed that some hematological malignancies, such as chronic lymphocytic leukemia, mantle cell lymphoma, may occur partly due to the constitutive activation of Wnt canonical signaling pathway. This review will summarize the latest development in Wnt canonical signaling pathway and its roles in tumorigenesis and angiogenesis.

Introduction

Wnt canonical signaling pathway acts a significant part in embryonic development and in maintenance of organs and tissues in adults. In the past two decades, medical scientists have devoted themselves to understanding the cellular and molecular mechanisms of Wnt signaling. A lot of studies indicate that Wnt canonical pathway involves in the pathogenesis of a range of disease including many kinds of carcinomas. Hematological malignancies are the types of carcinoma that affect blood, bone marrow and lymph nodes. They may derive from either of the two major blood cell lineages: myeloid and lymphoid cell lines. The incidence of hematological malignancies has been increasing steadily in the world for the past years, but their etiology and pathogenesis has not been well understood involving areas of chromosome aberrations, apoptosis inhibition, abnormal activation of signaling pathways, angiogenesis, et al. In this review, we focus on the role of Wnt canonical signaling in carcinomas, especially in hematological malignancies, and then disclose potential therapeutic opportunities of this pathway in hematological malignancies.

Wnt canonical pathway

Wnt signaling pathways are categorized as "canonical" and "non-canonical" Wnt pathways, which are β-catenin-

members in diverse species ranging from round worm and insects to human [1]. In the absence of a Wnt ligand binding to its receptor complex, the cytoplasmic β -catenin is degraded by the "destruction complex". In this complex, Axin acts as an scaffold protein, which adenomatous polyposis coli (APC), glycogen synthase kinase 3β (GSK-3β) and casein kinase 1α (CK1 α) bind to facilitate the sequential phophorylation of β-catenin in 45serine by kinase CK1 α and 41'threonine, 37',33'serine by GSK-3 β [2,3]. Accordingly, phosphorylated β-catenin is recognized by β-transducin-repeat-containing protein (β-TrCP) and constantly degraded by the ubiquitin-proteasome pathway. Wnt signaling is activated via ligation of Wnts to their respective dimeric cell surface receptors composed of the seven transmembrane frizzled (Fz) proteins and the lowdensity lipoprotein receptor-related protein 5/6 (LRP5/6). Upon ligation to their receptors, the cytoplasmic protein disheveled (Dvl) is recruited, phosphorylated and activated. Activation of Dvl induces the dissociation of GSK-3\beta from Axin and leads to the inhibition of GSK-3β. Next, the phosphorylation and degradation of β -catenin is inhibited as a result of the inactivation of the "destruction complex".

Subsequently, stabilized β -catenin translocates into the

nucleus. Nuclear β -catenin is the ultimate effector, binding

dependent and β-catenin-independent signaling pathways,

respectively. Here we will emphatically point out the role

of Wnt canonical pathway in hematological malignancies.

A simplified model of Wnt canonical pathway is deli-

neated in Fig. 1. Wnts is a group of secreted cysteine-rich

glycoproteins, which includes at least 19 identified



^{*} Correspondence: xinwang55@yahoo.com.cn Department of Hematology, Provincial Hospital Affiliated to Shandong University, Jinan, Shandong, 250021, China

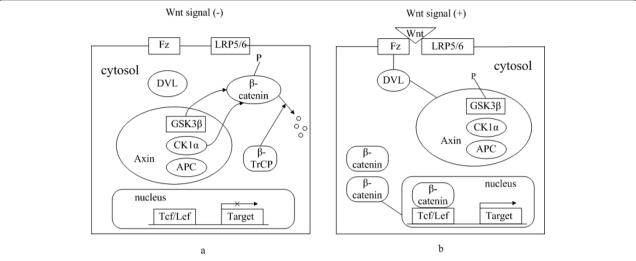


Figure 1 Wnt canonical pathway. (a) In the absence of a Wnt ligand, the cytoplasmic β -catenin is degraded by the "destruction complex". In this complex, Axin acts as an scaffold protein, which APC, GSK-3 β and CK1 α bind to facilitate the sequential phophorylation of β -catenin by kinase CK1 α and GSK-3 β . Accordingly, phosphorylated β -catenin is recognized by β -TrCP and constantly degraded by the ubiquitin-proteasome pathway. (b) Upon ligation of Wnts to their receptors composed of Fz proteins and LRP5/ δ , the cytoplasmic protein DvI is recruited, phosphorylated and activated. Activation of DvI induces the dissociation of GSK-3 β from Axin and leads to the inhibition of GSK-3 β . Next, the phosphorylation and degradation of β -catenin is inhibited as a result of the inactivation of the "destruction complex". Subsequently, stabilized β -catenin translocates into the nucleus. Nuclear β -catenin is the ultimate effector, binding to Tcf/Lef transcription factors to lead to changes in different target gene expressions.

to Tcf/Lef (T cell factor and lymphoid-enhancing factor) transcription factors that lead to changes in different target gene expressions that regulate cell proliferation, differentiation and survival, cell polarity and even angiogenesis.

Role of Wnt canonical signaling in carcinomas

Wnt canonical signaling is involved in pathogenesis of several carcinomas and the mechanisms of its over-activation are varied. Dysregulation of Wnt/β-catenin signaling plays a central role in early events in colorectal carcinogenesis. The APC protein which acts as a tumor suppressor protein can down-regulate the transcriptional activation mediated by Wnt/β-catenin. Therefore, inactivation of APC tumor suppressor gene caused by mutation is related to the initiation of colorectal neoplasia and its protein products lose the function of down-regulation of Wnt signaling. Then, colorectal cancer occurs. Furthermore, mutations of β -catenin in the functionally significant phosphorylation sites have been detected in colorectal tumors [4]. In melanoma cell lines, abnormally high amounts and stabilization of β-catenin accompanied by mutations in β-catenin or alteration/ missing of APC have been detected. Thus, genetic defects that result in up-regulation of β-catenin may play a role in melanoma progression [5]. Wnt canonical pathway has been confirmed to be related to initiation, development, progression and skeletal metastasis of prostate cancer in both human cancers and mouse models. It may result from mutation or altered expression of components of this pathway such as β-catenin and APC, which have been found in some types of prostate tumors and cancer cells [6,7]. Therefore, Wnt/β-catenin provides an attractive target for developing therapeutics of prostate cancers. Canonical Wnt pathway participates in many physiologic events in embryogenesis and is involved in embryogenic development of the ovary [8]. It also has an impact upon ovarian tumorigenesis especially a histologic subtype of epithelial ovarian cancer [9,10]. Non-small cell lung cancer(NSCLC) is one of the most common human carcinomas with a poor prognosis. Recent studies have revealed that the Wnt-1 overexpression, resulting in an aberrant and stabilized βcatenin expression, is associated with the expression of tumor-associated Wnt-targets(c-Myc, CyclinD1, Matrix Metalloproteinase 7), tumor proliferation, angiogenesis and a poor prognosis factor in NSCLC [11]. In addition, alterations of canonical Wnt signaling pathway due to frequent mutations in β -catenin have been detected in a wide range of other tumors, including hepatocellular carcinomas [12] and Wilms' tumors [13]. Mutations in the scaffold protein Axin [14] have been verified in some malignancies. Besides these mutations in intracellular signaling components, several tumors display a missing of expression of the secreted Wnt antagonists sFRPs and WIF1 resulting from silencing by promoter hypermethylation [15].

Since aberrant activation of Wnt canonical signaling pathway is diversely involved in pathogenesis of carcinomas, there has been great interest in developing therapeutics that circumvent it either by inhibiting Wnt mediated transcription or by inactivating the target genes. In some carcinomas, Wnt canonical signaling pathway has become the potential therapeutic target. The stabilized nuclear β-catenin due to the aberrant activation of Wnt signaling is an attractive therapeutic target for human cancers. Non-steroid anti-inflammatory drugs(NSAIDs) can suppress the activity of β-catenin. These drugs may inhibit Wnt/β-catenin signaling at multiple levels, including induction of β-catenin degradation [16] and disruption of the Tcf/β-catenin complex [17]. In addition, the transcriptional activity of Wnt/βcatenin can be inhibited by quercetin, a famous antitumor agent, in SW480 cell lines and also in HEK293 cells transiently transfected with constitutively active mutant β -catenin gene. The inhibitory mechanism is due to the decreased nuclear β -catenin and Tcf-4 proteins [18]. The way in which nuclear β-catenin and Tcf proteins were decreased needs to be further studied. The Wnt-Frizzled interaction can be antagonized by several secreted proteins, including Dickkopf1 (Dkk1), Wnt inhibitory factor 1(WIF-1) and secreted frizzledrelated protein (sFRP) family members. Wnt/β-catenin pathway can become the potential therapeutic target of cancer achieved by expression of secreted antagonists of the pathway, such as Dkk1. In addition, small interference of RNA (siRNA) may eliminate components of Wnt/ β -catenin signaling and can also be used to block this signaling. Others include small molecule inhibitors which can interfere the formation of the Tcf/β-catenin complex [19] or disturb the interaction of β-catenin with other co-activators [20] and monoclonal antibodies targeting the upstream signaling components such as Wnts ligands [21,22] or frizzled receptors, et al. The fact is that Wnt/β-catenin signaling pathway has great therapeutic potential in carcinomas.

Canonical Wnt signaling and angiogenesis

Wnt signaling pathway has been observed to make a difference in vessel development and pathology and in survival and proliferation of primary endothelial cells. Several Wnt ligands have been demonstrated to be expressed in vascular endothelial cells in vitro, including Wnt-7a, Wnt-10b and in vascular smooth muscle cells including Wnt-5a [23]. In vivo, the fetal vessels of the placenta express Wnt-2 [24] and the blood vessels of the mouse embryonic yolk sac express Wnt-5a and Wnt-10b [25]. Other components of this pathway such as Fz receptors have been demonstrated to be expressed in cultured endothelial cells and vascular smooth muscle cells [26,27]. During human embryonic development, nuclear and/or cytoplasmic β -catenin can be detected in placental villus capillaries, fetal capillaries, arteries and

veins [28]. Furthermore, Wnt/ β -catenin signaling may promote proliferation and survival in human endothelial cells via the induction of known angiogenic regulators; such as Interleukin-8 which is another transcriptional target of canonical Wnt pathway [29].

Angoigenesis is essential for tumor growth and metastasis. Studies have revealed the close relationship between canonical Wnt signaling pathway and angiogenesis of carcinomas. β-catenin accumulation has been involved in angiogenesis in brain cancer. β-catenin is found in the cytoplasm and nucleus of endothelium in neovessels of rat N-ethyl-N-nitrosurea-induced gliomas [30] and in the neovascular endothelial cells of medulloblastomas and other tumors of central nervous system [31]. However, accumulation of β -catenin in the cytoplasm or nucleus is rarely seen in cells of the normal adult brain vasculature [32]. A role for Wnt/β-catenin signaling in the vasculature is further supported by the identification of Wnt target genes that encode angiogenic regulators. Vascular endothelial growth factor A (VEGF-A) is a potent and widely distributed angiogenic peptide and has confirmed to be associated with the tumor angiogenesis and a poor prognosis [33,34]. It is also a target of canonical Wnt/β-catenin signaling pathway [35]. Seven β-catenin/Tcf binding sites occur in the VEGF-A promoter [36]. A recent study on NSCLC has disclosed that the Wnt1 expression correlates with the intratumoral VEGF-A expression with the action of elevating the activity of Wnt/β-catenin pathway [10]. In the meantime, a significant proportion of human colorectal cancers have an activating mutation in Wnt/ β-catenin pathway resulting in the abnormal expression of VEGF [37].

It is believed that Wnt signaling pathway is vital for tumor neovascularization and is a great potential in blocking tumor invasion and metastasis. To further confirm the role of Wnt/ β -catenin signaling pathway in tumor angiogenesis and growth, Wnt antagonists WIF1-Fc and sFRP1-Fc were used to treat hepatocellular carcinoma tumors. They revealed that these two fusion proteins could inhibit Wnt signaling and exerted potent antineoplastic activity by increasing apoptosis of tumor cells and by impairing tumor vascularization; including reducing the microvessel density, decreasing expression of vascular endothelial growth factor and stromal cell-derived factor-1 [38].

Role of Wnt canonical pathway in hematological malignancies

Hematopoiesis is a continuous process in which stem/progenitor cells develop into mature blood cellular components. Wnt/ β -catenin signaling pathway has been shown to have an effect on controlling the proliferation, survival and differentiation of hematopoietic cells [39].

The gene products of the Wnt family, functioning as hematopoietic growth factors, may exhibit higher specificity for earlier progenitor cells [40]. Wnts have additionally been shown to participate in hematopoiesis in which Wnt-11 induced bone marrow cells to develop into a variety of different lymphoid cell types [41]. Wnt3a signaling not only provides proliferative stimuli such as for immature thymocytes, but also regulates cell fate decisions of HSC during hematopoiesis [42]. More recently, gain of function studies have demonstrated that constitutively activated β-catenin in hematopoietic stem cells blocks multilineage differentiation; including B cell differentiation at early stages, suggesting the importance of fine tuning of Wnt/β-catenin signaling pathway for normal B cell development and function [43,44]. Frizzled 9 knockout in mice leads to abnormal B-cell development [45]. Wnt signaling is required for thymocyte development [46] and plays a key role in the maintenance of stemness in mature memory CD8+T cells [47]. Constitutive activation of β -catenin promotes the expansion of multipotential HSCs [44,48]. However, the influence of Wnt/β-catenin pathway on mature B cells is not obvious because they do not express TCF/ LEF factors [43]. Excessive stimulation of the Wnt cascade may lead to transformation of HSCs [44,48] and is noticeable in the neoplasms of myeloid and lymphoid lineages. Thus any aberrant signaling through this pathway may have a negative influence on hematopoiesis and may involve in lymphomagenesis.

Aberration of Wnt pathway and the related proteins are detected in many hematological patients [49]. Activation of Wnt signaling pathway has been implicated in the pathogenesis of leukemia. More recently, β-catenin activation coupled with GSK3\beta inactivation, has been demonstrated in chronic myeloid leukemia(CML) in blast crisis and precursor B-cell acute lymphoblastic leukemia(ALL) [50]. The function of canonical Wnt pathway is epigenetically regulated by methylation of Wnt antagonists and has prognostic relevance in acute myeloid leukemia(AML) [51]. Secreted Frizzled-related protein genes (sFRPs), functioning as Wnt signalling antagonists, have been found to be downregulated or inactivated by promoter hypermethylation in ALL and AML [52]. In addition, small molecule inhibitors of Wnt signaling effectively induce apoptosis in AML cells. Consequently, targeting this pathway seems to be an innovative approach in the treatment of AML [53]. Studies have demonstrated that deregulation of Wnt signaling pathway plays a role in the pathogenesis of CML. However, \beta-catenin amino-terminal mutations are not observed or are very rare and therefore are not the underlying mechanism of activated Wnt signaling in CML [54]. There must be other mechanisms for deregulating canonical Wnt signaling in CML. Wnt signaling genes are also overexpressed and may be pathologically reactivated in other neoplastic transformation of mature B cells, such as chronic lymphocytic leukemia (B-CLL). Uncontrolled Wnt signaling may contribute to defects in apoptosis that characterizes this malignancy [55,56].

Epstein-Barr Virus (EBV) is consistently detected in the endemic form of Burkitt's lymphoma (BL). An increase in both free and total β-catenin was seen in EBV-infected BL cells compared to EBV-negative cells [57]. The involvement of Wnt/β-catenin pathway in cell-cycle regulation, proliferation and invasion contributing to enhanced proliferative and metastatic properties of multiple myeloma (MM), were documented [58]. Furthermore, β-catenin small interfering RNA treatment inhibited the growth of multiple myeloma tumors in a xenograft model. As a result, β -catenin is the attractive novel target for treating multiple myeloma and other hematologic malignancies with aberrant canonical Wnt signaling [59]. Aberration of Wnt canonical pathway (WCP) may exist in mantle cell lymphoma(MCL) and appears to promote tumorigenesis in MCL. MCL tumors and cell lines highly and consistently expressed Wnt3 and Wnt10. Then, βcatenin was localized to the nucleus and transcriptionally active in MCL cell lines examined and more than half of the MCL tumors showed nuclear localization of β-catenin by immunohistochemistry, which obviously correlated with the expression of the phosphorylated/inactive form of GSK-3β(pGSK-3β) [60]. Of the clinical parameters, continuous pGSK-3β status had a significant correlation with absolute lymphocyte count in blood and negative pGSK-3β expression was significantly correlated with a longer overall survival in MCL [61]. Frequent βcatenin overexpression and accumulation may play an important part in the development of cutaneous lymphomas and it's mechanisms may not be associated with exon 3 mutation but others [62]. Nuclear localization of β-catenin was detected in extranodal marginal zone lymphoma by immunohistochemistry [63]. Scientists have revealed that esearchethacrynic acid (EA) and the antifungal agent ciclopiroxolamine (cic) could inhibit Wnt/βcatenin signalling in the myeloma cell line OPM-2 and three lymphoma cell lines (OCI-LY8-LAM-53, SU-DHL-4 and Raji) in vitro and led to apoptosis and a significant decrease of viability in lymphoma and its cell lines [64]. The Hedgehog (Hh) inhibitor, cyclopamine, and the Wnt inhibitor, quercetin, could suppress the growth of a number of leukemia and lymphoma cells [65]. Therefore, there is great potential that Wnt/β-catenin pathway can act as a therapeutic target of lymphoma and myeloma.

Conclusion and future directions

Wnt canonical signaling pathway is not only involved in cell survival, differentiation, apoptosis and maintenance of homeostasis, but also related to the pathogenesis of many carcinomas and hematological malignancies. Moreover, Wnt/β -catenin pathway has been revealed to be associated with angiogenesis of tumors. Its aberration has been detected in leukemia, myeloma and lymphoma. Canonical Wnt signaling may act as a potentially useful therapeutic target for hematological malignancies. Ultimately, further investigation is needed to interfere with Wnt signaling which may lead to new anti-cancer therapies.

Acknowledgements

This study was supported by the grants from the Natural Science Foundation of Shandong Province, China (No. Y2007C053 and No. ZR2009CM059) and the Project of Scientific and Technological Development of Shandong Province, China (No. 2007GG10002008).

Authors' contributions

Both authors participated in drafting and editing the manuscript. Both authors read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

Received: 22 July 2010 Accepted: 15 September 2010 Published: 15 September 2010

References

- Sidow A: Diversification of the Wnt gene family on the ancestral lineage of vertebrates. Proc Natl Acad Sci USA 1992, 89:5098-5102.
- Rubinfeld B, Albert I, Porfiri E, Fiol C, Munemitsu S, Polakis P: Binding of GSK3beta to the APC-beta-catenin complex and regulation of complex assembly. Science 1996, 272:1023-1026.
- Peifer M: Beta-catenin as oncogene: the smoking gun. Science 1997, 275:1752-1753.
- Morin PJ, Sparks AB, Korinek V, Barker N, Clevers H, Vogelstein B, Kinzler KW: Activation of beta-catenin-Tcf signaling in colon cancer by mutations in beta-catenin or APC. Science 1997, 275:1787-1790.
- Rubinfeld B, Robbins P, El-Gamil M, Albert I, Porfiri E, Polakis P: Stabilization of beta-catenin by genetic defects in melanoma cell lines. Science 1997, 275:1752-1753.
- Chesire DR, Ewing CM, Sauvageot J, Bova GS, Isaacs WB: Detection and analysis of beta-catenin mutations in prostate cancer. Prostate 2000, 45:323-334.
- de la Taille A, Rubin MA, Chen MW, Vacherot F, de Medina SG, Burchardt M, Buttyan R, Chopin D: Beta-catenin-related anomalies in apoptosisresistant and hormone-refractory prostate cancer cells. Clin Cancer Res 2003. 9:1801-1807
- Jeays-Ward K, Hoyle C, Brennan J, Dandonneau M, Alldus G, Capel B, Swain A: Endothelial and steroidogenic cell migration are regulated by WNT4 in the developing mammalian gonad. *Development* 2003, 130:3663-3670.
- Wu R, Zhai Y, Fearon ER, Cho KR: Diverse mechanisms of betacatenin deregulation in ovarian endometrioid adenocarcinomas. Cancer Res 2001, 61:8247-8255.
- Rask K, Nilsson A, Brannstrom M, Carlsson P, Hellberg P, Janson PO, Hedin L, Sundfeldt K: Wnt-signalling pathway in ovarian epithelial tumours: increased expression of beta-catenin and GSK3beta. Br J Cancer 2003, 89:1298-1304.
- Huang CL, Liu D, Ishikawa S, Nakashima T, Nakashima N, Yokomise H, Kadota K, Ueno M: Wnt1 overexpression promotes tumour progression in non-small cell lung cancer. EUROPEAN JOURNAL OF CANCER 2008, 44:2680-2688.
- de La Coste A, Romagnolo B, Billuart P, Renard CA, Buendia MA, Soubrane O, Fabre M, Chelly J, Beldjord C, Kahn A, Perret C: Somatic mutations of the beta-catenin gene are frequent in mouse and human hepatocellular carcinomas. Proc Natl Acad Sci USA 1998, 95:8847-8851.

- 13. Maiti S, Alam R, Amos CI, Huff V: Frequent association of beta-catenin and WT1 mutations in Wilms tumors. *Cancer Res* 2000, **60**:6288-6292.
- Behrens J, Jerchow BA, Würtele M, Grimm J, Asbrand C, Wirtz R, Kühl M, Wedlich D, Birchmeier W: Functional interaction of an axin homolog, conductin, with beta-catenin, APC, and GSK3beta. Science 1998, 280:596-599.
- Mazieres J, He B, You L, Xu Z, Lee AY, Mikami I, Reguart N, Rosell R, McCormick F, Jablons DM: Wnt inhibitory factor-1 is silenced by promoter hypermethylation in human lung cancer. Cancer Res 2004, 64:4717-4720.
- Bos CL, Kodach LL, van den Brink GR, Diks SH, van Santen MM, Richel DJ, Peppelenbosch MP, Hardwick JC: Effect of aspirin on the Wnt/betacatenin pathway is mediated via protein phosphatase 2A. Oncogene 2006, 25:6447-6456.
- Nath N, Kashfi K, Chen J, Rigas B: Nitric oxide-donating aspirin inhibits beta-catenin/T cell factor (TCF) signaling in SW480 colon cancer cells by disrupting the nuclear beta-catenin-TCF association. Proc Natl Acad Sci USA 2003, 100:12584-12589.
- Park Hoon Chi, Chang Young Joon, Hahm Ryeong Eun, Park Seyeon, Kim Hyun-Kyung, Yang Hak Chul: Quercetin, a potent inhibitor against βcatenin/Tcf signaling in SW480 colon cancer cells. Biochemical and Biophysical Research Communications 2005, 328:227-234.
- Lepourcelet M, Chen YN, France DS, Wang H, Crews P, Petersen F, Bruseo C, Wood AW, Shivdasani RA: Small-molecule antagonists of the oncogenic Tcf/beta-catenin protein complex. Cancer Cell 2004, 5:91-102.
- Eguchi M, Nguyen C, Lee SC, Kahn M: ICG-001, a novel small molecule regulator of TCF/beta-catenin transcription. Med Chem 2005, 1:467-472.
- He B, Reguart N, You L, Mazieres J, Xu Z, Lee AY, Mikami I, McCormick F, Jablons DM: Blockade of Wnt-1 signaling induces apoptosis in human colorectal cancer cells containing downstream mutations. *Oncogene* 2005, 24:3054-3058.
- You L, He B, Xu Z, Uematsu K, Mazieres J, Fujii N, Mikami I, Reguart N, McIntosh JK, Kashani-Sabet M, McCormick F, Jablons DM: An anti-Wnt-2 monoclonal antibody induces apoptosis in malignant melanoma cells and inhibits tumor growth. Cancer Res 2004, 64:5385-5389.
- Wright M, Aikawa M, Szeto W, Papkoff J: Identification of a Wntresponsive signal transduction pathway in primary endothelial cells. Biochem Biophys Res Commun 1999, 263:384-388.
- Monkley SJ, Delaney SJ, Pennisi DJ, Christiansen JH, Wainwright BJ: Targeted disruption of the Wnt2 gene results in placentation defects. Development 1996, 122:3343-3353.
- Ishikawa T, Tamai Y, Zorn AM, Yoshida H, Seldin MF, Nishikawa S, Taketo MM: Mouse Wnt receptor gene Fzd5 is essential for yolk sac and placental angiogenesis. Development 2001, 128:25-33.
- Mao C, Malek OT, Pueyo ME, Steg PG, Soubrier F: Differential expression of rat frizzled-related frzb-1 and frizzled receptor fz1 and fz2 genes in the rat aorta after balloon injury. Arterioscler Thromb Vasc Biol 2000, 20:43-51.
- Sala CF, Formenti E, Terstappen GC, Caricasole A: Identification, gene structure, and expression of human frizzled-3 (FZD3). Biochem Biophys Res Commun 2000, 273:27-34.
- Eberhart CG, Argani P: Wnt signaling in human development: Betacatenin nuclear translocation in fetal lung, kidney, placenta, capillaries, adrenal, and cartilage. Pediatr Dev Pathol 2001, 4:351-357.
- Ne'stor T, Masckaucha'n H, Shawber JCarrie, Funahashi Yasuhiro, Li Chi-Ming, Kitajewski Jan: Wnt/β-catenin signaling induces proliferation, survival and Interleukin-8 in human endothelial cells. Angiogenesis 2005, 8:43-51.
- Yano H, Hara A, Shinoda J, Takenaka K, Yoshimi N, Mori H, Sakai N: Immunohistochemical analysis of beta-catenin in N-ethyl-N-nitrosoureainduced rat gliomas: Implications in regulation ofangiogenesis. Neurol Res 2000, 22:527-532.
- Eberhart CG, Tihan T, Burger PC: Nuclear localization and mutation of beta-catenin in medulloblastomas. J Neuropathol Exp Neurol 2000, 59:333-337.
- Yano H, Hara A, Takenaka K, Shinoda J, Shimokawa K, Yoshimi N, Mori H, Sakai N: Differential expression of beta-catenin in human glioblastoma multiforme and normal brain tissue. Neurol Res 2000, 22:650-656.
- Huang C, Liu D, Masuya D, Nakashima T, Kameyama K, Ishikawa S, Ueno M, Haba R, Yokomise H: Clinical application of biological markers for treatments of resectable non-small-cell lung cancers. Br J Cancer 2005, 92:1231-1239.

- Gratzinger D, Zhao S, Marinelli RJ, Kapp AV, Tibshirani RJ, Hammer AS, Hamilton-Dutoit S, Natkunam Y: Microvessel Density and Expression of Vascular Endothelial Growth Factor and Its Receptors in Diffuse Large B-Cell Lymphoma Subtypes. The American Journal of Pathology 2007, 170:1362-1369.
- Zhang X, Gaspard JP, Chung DC: Regulation of vascular endothelial growth factor by the Wnt and K-ras pathways in colonic neoplasia. Cancer Res 2001, 61:6050-6054.
- Easwaran V, Lee SH, Inge L, Guo L, Goldbeck C, Garrett E, Wiesmann M, Garcia PD, Fuller JH, Chan V, Randazzo F, Gundel R, Warren RS, Escobedo J, Aukerman SL, Taylor RN, Fantl WJ: beta-Catenin regulates vascular endothelial growth factor expression in colon cancer. Cancer Res 2003, 63:3145-3153.
- 37. Huang D, Du X: Crosstalk between tumor cells and microenvironment via wnt pathway in colorectal cancer dissemination. *World J Gastroenterol* 2008, **14**:1823-1827.
- Hu J, Dong A, Fernandez-Ruiz V, Shan J, Kawa M, Martínez-Ansó E, Prieto J, Qian C: Blockade of Wnt signaling inhibits angiogenesis and tumor growth in hepatocellular carcinoma. Cancer Res 2009, 69:6951-6959.
- Reya T, Duncan AW, Ailles L, Domen J, Scherer DC, Willert K, Hintz L, Nusse R, Weissman IL: A role for Wnt signalling in self-renewal of haematopoietic stem cells. Nature 2003, 423:409-414.
- Van Den Berg JDavid, Sharma KArun, Bruno Edward, Hoffman Ron: Role of Members of the Wnt Gene Family in Human Hematopoiesis. Blood 1998, 92:3189-3202
- 41. Brandon C, Eisenberg LM, Eisenberg CA: WNT signaling modulates the diversification of hematopoietic cells. *Blood* 2000, 96:4132-4141.
- Luis TC, Weerkamp F, Naber BA, Baert MR, de Haas EF, Nikolic T, Heuvelmans S, De Krijger RR, van Dongen JJ, Staal FJ: Wnt3a deficiency irreversibly impairs hematopoietic stem cell self-renewal and leads to defects in progenitor cell differentiation. *Blood* 2009, 113:546-554.
- Reya T, O'Riordan M, Okamura R, Devaney E, Willert K, Nusse R, Grosschedl R: Wnt signaling regulates B lymphocyte proliferation through a LEF-1 dependent mechanism. *Immunity* 2000, 13:15-24.
- Baba Y, Yokota T, Spits H, Garrett KP, Hayashi S, Kincade PW: Constitutively active beta-catenin promotes expansion of multipotent hematopoietic progenitors in culture. J. Immunol 2006, 177:2294-2303.
- Ranheim AErik, Kwan CKHelen, Reya Tannishtha, Wang Yu-Ker, Weissman Lirving, Francke Uta: Frizzled 9 knock-out mice have abnormal B-cell development. Blood 2005, 105:2487-2494.
- Staal FJ, Meeldijk J, Moerer P, Jay P, van de Weerdt BC, Vainio S, Nolan GP, Clevers H: Wnt signaling is required for thymocyte development and activates Tcf-1 mediated transcription. Eur J Immunol 2001. 31:285-293.
- Gattinoni L, Zhong XS, Palmer DC, Ji Y, Hinrichs CS, Yu Z, Wrzesinski C, Boni A, Cassard L, Garvin LM, Paulos CM, Muranski P, Restifo NP: Wnt signaling arrests effector T cell differentiation and generates CD8+ memory stem cells. Nat Med 2009, 15:808-813.
- Baba Y, Garrett KP, Kincade PW: Constitutively active beta-catenin confers multilineage differentiation potential on lymphoid and myeloid progenitors. *Immunity* 2005, 23:599-609.
- Weerkamp F, van Dongen JJ, Staal FJ: Notch and Wnt signaling in T-lymphocyte development and acute lymphoblastic leukemia. *Leukemia* 2006, 20:1197-1205.
- Khan NI, Bradstock KF, Bendall LJ: Activation of Wnt/beta-catenin pathway mediates growth and survival in B-cell progenitor acute lymphoblastic leukaemia. Br J Haematol 2007, 138:338-348.
- Valencia A, Román-Gómez J, Cervera J, Such E, Barragán E, Bolufer P, Moscardó F, Sanz GF, Sanz MA: Wnt signaling pathway is epigenetically regulated by methylation of Wnt antagonists in acute myeloid leukemia. Leukemia 2009, 23:1658-1666.
- Jost E, Schmid J, Wilop S, Schubert C, Suzuki H, Herman JG, Osieka R, Galm O: Epigenetic inactivation of secreted Frizzled-related proteins in acute myeloid leukaemia. Br J Haematol 2008, 142:745-753.
- Minke KS, Staib P, Puetter A, Gehrke I, Gandhirajan RK, Schlösser A, Schmitt EK, Hallek M, Kreuzer KA: Small molecule inhibitors of WNT signaling effectively induce apoptosis in acute myeloid leukemia cells. Eur J Haematol 2009. 82:165-175.
- Sercan Z, Pehlivan M, Gokturk D, Sercan HO: Beta-catenin mutations are not observed in chronic myeloid leukemia. *Tumori* 2009, 95:836-839.

- Lu D, Zhao Y, Tawatao R, Cottam HB, Sen M, Leoni LM, Kipps TJ, Corr M, Carson DA: Activation of the Wnt signaling pathway in chronic lymphocytic leukemia. PNAS 2004, 101:3118-3123.
- Wu QL, Zierold C, Ranheim EA: Dysregulation of Frizzled 6 is a critical component of B-cell leukemogenesis in a mouse model of chronic lymphocytic leukemia. *Blood* 2009, 113:3031-3039.
- Éverly DN Jr, Kusano S, Raab-Traub N: Accumulation of cytoplasmic betacatenin and nuclear glycogen synthase kinase 3beta in Epstein-Barr virus-infected cells. J Virol 2004, 78:11648-11655.
- Dutta-Simmons J, Zhang Y, Gorgun G, Gatt M, Mani M, Hideshima T, Takada K, Carlson NE, Carrasco DE, Tai YT, Raje N, Letai AG, Anderson KC, Carrasco DR: Aurora kinase A is a target of Wnt/beta-catenin involved in multiple myeloma disease progression. *Blood* 2009, 114:2699-2708.
- Ashihara E, Kawata E, Nakagawa Y, Shimazaski C, Kuroda J, Taniguchi K, Uchiyama H, Tanaka R, Yokota A, Takeuchi M, Kamitsuji Y, Inaba T, Taniwaki M, Kimura S, Maekawa T: beta-catenin small interfering RNA successfully suppressed progression of multiple myeloma in a mouse model. Clin Cancer Res 2009, 15:2731-2738.
- Gelebart P, Anand M, Armanious H, Peters AC, Dien Bard J, Amin HM, Lai R: Constitutive activation of the Wnt canonical pathway in mantle cell lymphoma. *Blood* 2008, 112:5171-5179.
- 61. Chung R, Peters AC, Armanious H, Anand M, Gelebart P, Lai R: Biological and clinical significance of GSK-3beta in mantle cell lymphoma–an immunohistochemical study. *Int J Clin Exp Pathol* 2010, **3**:244-253.
- Bellei Barbara, Pacchiarotti Alberto, Perez Marie, Faraggiana Tullio: Frequent b-catenin overexpression without exon 3 mutation in cutaneous lymphomas. Modern Pathology 2004, 17:1275-1281.
- Zhang D, O'neil MF, Cunningham MT, Fan F, Olyaee M, Li L: Abnormal Wnt signaling and stem cell activation in reactive lymphoid tissue and lowgrade marginal zone lymphoma. *Leuk Lymphoma* 2010, 51:906-910.
- Schmidt Matthias, Sievers Elisabeth, Endo Tomoyuki, Desheng Lu, Carson Dennis, Schmidt-Wolf GHIngo: Targeting Wnt pathway in lymphoma and myeloma cells. British Journal of Haematology 2008, 144:796-798.
- Kawahara T, Kawaguchi-Ihara N, Okuhashi Y, Itoh M, Nara N, Tohda S: Cyclopamine and quercetin suppress the growth of leukemia and lymphoma cells. Anticancer Res 2009, 29:4629-4632.

doi:10.1186/1756-8722-3-33

Cite this article as: GE and Wang: Role of Wnt canonical pathway in hematological malignancies. *Journal of Hematology & Oncology* 2010 **3**:33.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at www.biomedcentral.com/submit

