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ABT-869, a promising multi-targeted tyrosine kinase inhibitor: from bench to bedside

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Abstract

Tyrosine Kinase Inhibitors (TKI) have significantly changed the landscape of current cancer therapy. Understanding of mechanisms of aberrant TK signaling and strategies to inhibit TKs in cancer, further promote the development of novel agents.

ABT-869, a novel ATP-competitive receptor tyrosine kinase inhibitor is a potent inhibitor of members of the vascular endothelial growth factor (VEGF) and platelet derived growth factor (PDGF) receptor families. ABT-869 showed potent antiproliferative and apoptotic properties in vitro and in animal cancer xenograft models using tumor cell lines that were "addicted" to signaling of kinases targeted by ABT-869. When given together with chemotherapy or mTOR inhibitors, ABT-869 showed at least additive therapeutic effects. The phase I trial for ABT-869 was recently completed and it demonstrated respectable efficacy in solid tumors including lung and hepatocellular carcinoma with manageable side effects. Tumor cavitation and reduction of contrast enhancement after ABT-869 treatment supported the antiangiogenic activity. The correlative laboratory studies conducted with the trial also highlight potential biomarkers for future patient selection and treatment outcome.

Parallel to the clinical development, *in vitro* studies on ABT-869 resistance phenotype identified novel resistance mechanism that may be applicable to other TKIs. The future therapeutic roles of ABT-869 are currently been tested in phase II trials.

Introduction

Receptor tyrosine kinases (RTKs) and protein phosphatases control reversible protein phosphorylation [1,2]. This process mediates critical signaling transduction between cell and extracellular stimulation, including survival, growth and differentiation. Dysregulation of RTK

signaling pathways has been correlated with the progression of cancers with different histological origins [1]. For example, amplification of the HER2 gene is observed in ~30% of breast cancer biopsies and forms the basis for the use of trastuzumab (Herceptin, Genentech, Inc, California) to treat breast cancer patients.

The common molecular mechanisms underlying such aberrant activities are point mutation, duplication, and amplification of the RTK, which leads to gain-of-function and consecutive activation of the kinases in general. The fms-like tyrosine kinase 3 (FLT3) is a class III RTK family and shares strong structural similarity with other family members including receptors for platelet-derived growth factors A (PDGFRA) and B (PDGFRB), the colony-stimulating factor 1 receptor (CSF1-R) and steel factor receptor (KIT) [3-5]. FLT3 mutations are identified in about onethird of adult acute myeloid leukemia (AML) [6-10]. The interactions between the vascular endothelial growth factors (VEGF) and their receptors (VEGFRs) are crucial for angiogenesis [11,12]. The expression of VEGF and its receptors are detected in most of solid tumors and hematological malignancies [13]. Overexpression of VEGF and/ or it's receptor VEGFR2 contributes to invasiveness and metastasis of breast, lung, prostate, renal-cell, colon cancers and hepatocellular carcinoma [11,12]. In AML, a number of studies have demonstrated that an autocrine/ paracrine pathway between VEGF and its receptors are involved in poor survival of a subset of patients and progression of the disease [14-17]. This evidence underpins an important discovery in the molecular biology of cancer that histological different types of cancer could share the same dysregulated signaling pathway(s) and one particular type of cancer could have multiple genetic abnormalities. Therefore, there has been great interest in discovering compounds targeting multiple RTKs with the rationale of potential superior antitumor activity for a variety of cancer types.

ABT-869, a novel ATP-competitive RTK inhibitor, is active against all VEGFRs and PDGFR families, but minimally active against unrelated RTKs and cytosolic tyrosine kinases and serine/threonine kinases [18]. The goals of this article are to summarize the published data on preclinical and clinical development of ABT-869, an orally active multi-targeted RTK inhibitor in the treatment of leukemia and solid tumors. Secondly, various strategies and rationale as well as mechanistic studies of combining ABT-869 with other agents will be reviewed. Lastly, we discuss the potential drug resistance issue in ABT-869 therapy based on our laboratory's published data. ABT-869 is under active clinical development primarily in solid tumors and early phase data and ongoing phase II studies will be reviewed.

The chemical structure and target selection of ABT-869

ABT-869 was discovered in Abbott Laboratories (Abbott Park, IL, USA) through a structure-based rational design, by incorporating an N, N'-diaryl urea moiety at the C4-position of 3-aminodazole (Figure 1) [19]. The molecular weight of ABT-869 is 375.4. ABT-869 shows potent effi-

Figure I
The chemical structure of ABT-869: N- [4-(3-amino-IH-indazol-4-yl)phenyl]- NI-(2-fluoro-5-methylphenyl) urea.

cacy to inhibit all the members of VEGFR and PDGFR family with nanomolar range of IC₅₀, but much less activity to other nonrelated tyrosine kinase (Table 1) [18]. The selectivity profile of ABT-869 against a broader range of kinases is illustrated in Figure 2. In comparison to 5 other multitargeted RTK inhibitors (PTK787 [Vatalanib*, Novartis-Schering AG], AG013736 [Axitinib*, Pfizer], BAY43-9006 [Nexavar*, Bayer], CHIR258 [Chiron], and SU11248 [Sutent*, Pfizer]) [19], that have undergone clinical development, ABT-869 inhibited a broader number of kinases relevant to the VEGF signaling pathway. AG013736, CHIR258, and SU11248 are also active against most of the targeted kinases but these inhibitors demonstrate more off-target activity than ABT-869 [18].

Another potentially important aspect of the distinctive activity profile of ABT-869 is the molecule's activity against CSF1R [20]. This activity is manifested as potent inhibition of CSF-1R signaling in macrophage-derived cells [21]. In vivo activity of ABT-869 for inhibiting CSF1R-mediated responses is exemplified by results illustrated in Figure 3 showing the effect of oral administration of ABT-869 on CSF1 priming of LPS-induced TNF release in mice. This activity may contribute to the anti-tumor activity of ABT-869 in cancer models where elevated levels of inflammatory tumor-associated macrophages drive tumor progression.

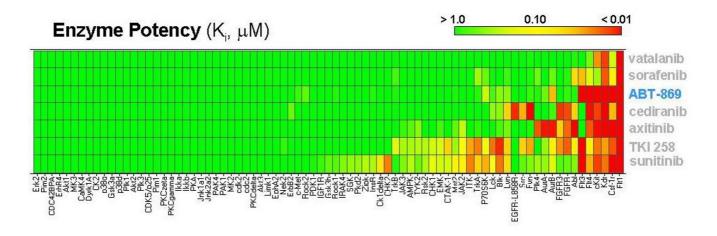


Figure 2
Kinase inhibition profile of ABT-869 against a broader range of kinases.

Nonclinical in vivo activity of ABT-869

Initial nonclinical studies demonstrated potent antiproliferative and apoptotic effects of ABT-869 on cancer cells whose proliferation is dependent on mutant kinases, such as FLT3 [18,20,22]. ABT-869 given orally was effective in multiple in vivo human xenograft tumor growth models and showed *in vivo* mechanism-based targeting, including acute myeloid leukemia with FLT3 mutation (MV4–11), highly angiogenic fibrosarcoma (HT1080), small cell lung carcinoma (H526, known to express KIT), colon adenocarcinoma (DLD-1), epidermoid carcinoma (A431) and breast cancinoma (MX-1). In addition to flank xenografts, ABT-869 has demonstrated dose dependant efficacy in orthotopic tumor growth models with the breast carci-

Table 1: Kinase inhibition profile of ABT-869 (with permission adapted from Molecular Cancer Therapeutics 2006;5:995–1006)

Related RTK ^a		Non-related TK ^a		Ser/Thr Kinases ^b		
Kinase	IC ₅₀ (nM)	Kinase	IC ₅₀ (nM)	Kinase	IC ₅₀ (nM)	
KDR	8	SRC	> 50,000	AKT	> 50,000	
FLTI	3	IGFR	> 50,000	SGK	940	
FLT4	40	INSR	> 50,000	CDC2	9,800	
$PDGFR\alpha$	29	LCK	38,000	PKA	5,900	
$PDGFR\beta$	25	EGFR	> 50,000			
CSF-1R	5	HCK	> 50,000			
KIT	20	CMET	> 50,000			
FLT3	10	LYN	> 20,000			
TIE2	170	FYN	> 50,000			
RET	1,900	FGR	> 50,000			
FGFR	> 12,500					

a. IC_{50} values determined at an ATP concentration of 1 mM.

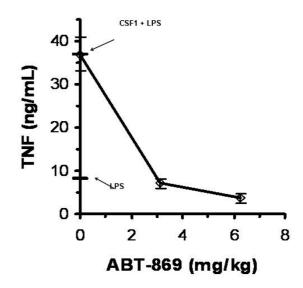


Figure 3 Inhibition of CSF1-primed LPS-induced TNF release. Mice were given ABT-869 (PO) at the indicated dose and 45 minutes later primed with CSF1 (1.8 μ g IP). After 3.25 hours, LPS (300 μ g IP) was administered. Serum TNF, expressed as mean \pm SEM (n = 6), was assessed 1.5 hours later. CSF1 increased serum TNF induced by LPS by >4 fold (8 vs 37 ng/mL).

b. IC_{50}^{30} values determined at an ATP concentration of 5 to 10 $\mu\text{M}.$

noma cell lines MDA-231 (epithelial) and MDA-435LM (ductal) as well as a rat glioma cell line (9L). ABT-869 was also efficacious at inhibiting the growth of prostate cancer cells in a bone environment, thereby demonstrating potential therapeutic utility in a metastases setting [23]. A summary of activity in these and other tumor models is presented in Figure 4.

In addition to single agent activity ABT-869 also exhibited antitumor activity when given in combination with chemotherapy agents, including: carboplatin, cisplatin, docetaxel, gemcitabine, irinotecan, paclitaxel, rapamycin, TMZ and Ara-C [18,22,24,25]. The effect of combination therapy with carboplatin-paclitaxel (dosed concurrently) on the dose-dependent activity of ABT-869 in a NSCLC model response is shown in Figure 5. This response to combination therapy is typical in that it reflects an increase in efficacy with no increase in overall toxicity. However, the outcome of combination therapy can be somewhat sequence-dependent, as is discussed below.

In light of its preclinical activity profile, ABT-869 underwent the industrial standard pre-clinical toxicology, metabolism, and pharmacology studies and the compound was deemed to be suitable to further clinical development (see below).

Nonclinical studies of ABT-869 and in combination with chemotherapy in acute myeloid leukemia with and without FLT-3 mutations

Approximately, 25% of AML patients have acquired FLT3 internal tandem duplications (FLT3-ITDs), varying from 3 to \geq 400 base pairs in the juxtamembrane domain, and 7% of AML patients harbor activating point mutations in the second kinase domain (FLT3-TK) [7-10]. FLT3 mutations therefore represent the most common genetic alteration in AML and therefore, have been targeted for therapeutic agent development. Patents with FLT3-ITD are usually associated with poor outcome, but the prognosis of FLT3-TK mutation remains inconclusive [7-10]. FLT3-ITD mutations trigger strong autophosphorylation of the FLT3 kinase domain, and constitutively activate several downstream effectors such as the PI3K/AKT pathway, RAS/MAPK pathway, and the STAT pathway, mainly STAT5 (Figure 6). Oncogenic protein kinase PIM1 also is up-regulated by FLT3-ITD. These rampant signaling pathways are wired to promote uncontrolled cell survival and proliferation, leading to transformation of leukemia [26].

For leukemia cell lines with FLT3-ITD such as MV4–11 and MOLM-14, ABT-869 potently inhibits their proliferation at IC $_{50}$ less than 10 nM [22,27]. ABT-869 also induces dose-dependently $\rm G_1$ cell cycle arrest and apoptosis in these FLT3-ITD positive cells [22,27]. Analysis of key cell

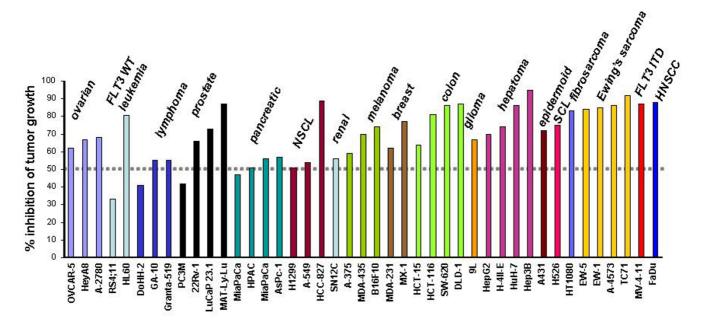


Figure 4
Efficacy of ABT-869 in representative xenografts. Efficacy was defined as percent of tumor size relative to vehicle-treated remaining after 3–4 weeks of dosing ABT-869 (10–25 mg/kg/day).

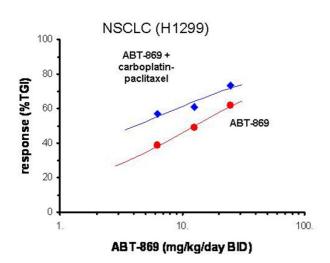


Figure 5
Efficacy of ABT-869 in combination with carboplatinpaclitaxel in a NSCLC xenograft. ABT-869 was administered orally at the indicated dose for 3 weeks and carboplatin-paclitaxel was administered weekly (IP and IV
respectively) beginning 3 weeks after inoculation of H1299
cells into the flank of SCID/beige mice. Percent inhibition of
tumor size relative to vehicle treated control was calculated
at the end of the study is indicated in parentheses in the legend.

cycle regulators reveals that simultaneous terminal reduction of cyclins D and E, the key G_1/S cyclins, and progressive increases in cyclin dependent kinase inhibitors (CDKIs) p21^{waf1/Cip}, p27^{kip1} contributed to the blockage of G_1/S progression induced by ABT-869 [22]. ABT-869 increases the expression of a few proapoptotic proteins including BAD, BAK and BID, and decreases the pro-survival molecule Bcl-xL. Cleaved BID and PARP, a hallmark of apoptosis, is evident [22].

ABT-869, as predicted from its kinase inhibition profile, targets the FLT3 signaling pathway. In MV4–11 cells, ABT-869 inhibits phosphorylation of FLT3 receptor (p-FLT3), as well as downstream signaling effectors p-AKT, p-ERK, p-STAT5 and PIM-1 kinase at a concentration of 1 nM [22,27]. Importantly, ABT-869 has been shown to effectively inhibit colony formation of primary AML bone marrow cells at 100 nM, but no inhibition on normal human bone marrow progenitor cells up to 1 μ M, suggesting ABT-869 is not toxic to normal bone marrow cells [27]. In a mice bone marrow engraftment model of MV4–11 cells, ABT-869 treatment significantly prolonged survival and reduced leukemic burden (CD45+ human cells)

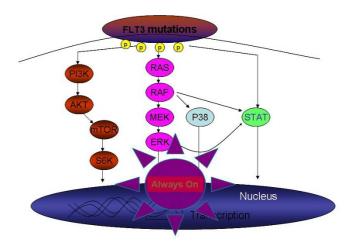


Figure 6
The FLT3-ITD signaling pathways. The presence of FLT3-ITD induces ligand-independent receptor dimerization and activates three major signaling pathways including PI3K/AKT, MAPK and STAT5 pathways. These signalings are transferred to nucleus, which lead to the transcription of genes involved in cell proliferation and survival.

in a dose-dependent fashion when compared to vehicle control treatment [27].

However, considering the complexity of the disease, ABT-869 as a single agent is unlikely to deliver complete or lasting responses in AML. We demonstrated that ABT-869 also produces synergistic antileukemic effect with chemotherapy in a sequence dependent manner [22]. This sequence-specific synergism was also demonstrated with another FLT3 inhibitor, CEP-701 (Lestaurtinib[®], Cephalon, Inc., Frazer, PA, USA) [28]. For simultaneous treatment in MV4-11 and MOLM-14 cells, combination of lower doses of ABT-869 and cytosine arabinoside (Ara-C) generates an additive or mildly synergistic interaction. All of the combinations of ABT-869 and Doxorubicin (Dox) results in synergistic effects. However, pretreatment with ABT-869 antagonizes the cytotoxicity of Ara-C and Dox [22]. In contrast, chemotherapy (either Ara-C or Dox) followed by ABT-869 produces significant synergism on inhibition of proliferation and induction of apoptosis in MV4-11 and MOLM-14 cells, as well as primary patient AML cells with FLT3-ITD mutations [22]. In a MV4-11 tumor xenograft model, combination of Ara-C at 15 mg/kg/day for 4 days and ABT-869 at 15 mg/kg/day results in faster reduction of tumor burden compared to ABT-869 treatment alone. Importantly, no adverse side effect is observed in the combination treatment group in terms of behavior or body weight changes [22]. Low density array (LDA) analysis reveals that inhibition of cell cycle related genes and MAPK pathway play an important role in the synergistic mechanism. Particularly, Cyclin D1

(CCND1) and Moloney murine sarcoma viral oncogene homolog (c-Mos) were the two most significantly down-regulated genes [22]. Collectively, these studies help to define the optimal combination sequence of chemotherapy and ABT-869 for clinical trials in AML.

Neoangiogenesis plays an important role in the pathogenesis of AML, so targeting VEGF/VEGFR receptors appears to be an alternative approach for treating AML [13]. Based on the early promising clinical trial results in AML patients regardless of FLT3 status achieved by other multitargeted inhibitors like SU11248 and PTK787/ZK 222584 [29-31]. ABT-869 was also tested against a wild type FLT3-AML cell line, HL60 in a xenograft model. HL60-RFP, a stable transfectant with red fluorescence protein, was examined in both the subcutaneous and systemic leukemia xenograft models using an advanced Olympus OV100 Whole-Animal Imaging System [32]. ABT-869 reduces leukemia burden and prolongs survival of NOD/ SCID mice engrafted with HL60-RFP. ABT-869 is effective in delaying tumor growth about five-fold in the subcutaneous xenograft model (Figure 7) by inhibiting angiogenesis via VEGF/VEGFRs loop [32].

Nonclinical studies of ABT-869 as a single agent and in combination with mTOR inhibitor in Hepatocellular carcinoma (HCC)

Expression of VEGF, the primary pro-angiogenic factor, has higher in HCC than in normal hepatic parenchyma cells and has been shown to positively correlate with vascularization of HCC [33,34]. HCC cells are dependent on the supply of oxygen and nutrient through this neoangiogenesis [33,34]. Consequently, inhibition of neoangiogenesis could serve as a promising approach for the intervention of HCC.

In addition, the mammalian target of rapamycin (mTOR), a cytosolic serine/threonine kinase, has emerged as an attractive anticancer target in recent years [35]. mTOR plays an essential role not only in controlling the mammalian translation machinery, but also in regulating signaling pathways that respond to growth factors and nutrition. Activation of mTOR enhances translation of mRNAs that encodes key regulation protein for cell cycle, cell proliferation and growth such as cyclin D148 and ornithine decarboxylase 49 by phosphorylation of S6K1 (p70S6 kinase) and 4E-BP1 (EIF4-binding protein 1) [36]. mTOR is also a central downstream effector of PI3K/AKT pathways.[37] The mTOR signaling pathway has been reported to be deregulated in HCC [38,39]. Rapamycin, a mTOR inhibitor, binds to the immunophilin FKBP12, and the formed complex inactivates mTOR, further suppressing p70S6 kinase and 4E-BP1, two critical downstream targets of mTOR signaling. Rapamycin inhibits proliferation of HCC cell lines, including HepG2, Hep3B,

and Sk-hep-1 [40,41]. Therefore, combining ABT-869 with rapamycin would be a reasonable targeted therapy for HCC.

We demonstrated that oral administration of ABT-869 as a single agent at a dose of 10 mg/kg/day effectively inhibits the growth of Huh7 and Sk-hep-1 tumors in mouse xenograft models [24]. ABT-869 shows a dramatic inhibition of neoangiogenesis in vivo. This is supported by immunohistochemistry (IHC) analysis that shows ABT-869 significantly down-regulates VEGF and reduces the formation of Microvessel density (MVD). Bevacizumab, a specific anti-VEGF antibody, was also compared with ABT-869 in a Sk-hep-1 mouse xenograft. The antitumor activity of ABT-869 is significantly higher than bevacizumab in this model [24]. Further analysis reveals that phosphorylation of p44/42 MAP kinase is also substantially decreased in the ABT-869-treated tumor samples [24]. The additional targeting achieved by the multi-targeted properties of ABT-869 could explain the significant advantage of anti-angiogenic activity of ABT-869 over bevacizumab, since MAPK pathway is known to be dsyregulated in human HCC.

Combination of ABT-869 (10 mg/kg/day) with Rapamycin (2 mg/kg/day) shows significant tumor volume reduction in both Huh7 and Sk-hep-1 animal models when compared to either of the single drug treatments (p < 0.05). Up-regulation of the cell cycle inhibitor, p27, and inhibition of the MAPK pathway contribute to the synergistic antitumor effect observed in combination therapy [24]

Taken together, these results support the rationale for clinical development of combination therapy of ABT-869 and other chemotherapies such as Rapamycin in HCC.

Dissecting the potential resistance phenomenon in ABT-869

In contrast to their potent efficacy in cellular based assays and xenograft models, in clinical trials, FLT3 inhibitors alone only achieve moderate and transient responses in the majority of AML patients [29,42-45]. Furthermore, important experience has been gained from imatinib mesylate (Gleevec) used as monotherapy for treating chronic myeloid leukemia (CML) indicating that under prolonged therapy with TKIs, patients could develop resistance or relapse [46]. Point mutations in the ATP binding site or gene amplification of BCR-ABL are the main cause of imatinib-resistance in CML patients [47]. However, point mutations in the FLT3 kinase domain are not common [48,49].

As ABT-869 was entering early phase clinical development with continuous daily dosing schedule, we investigated

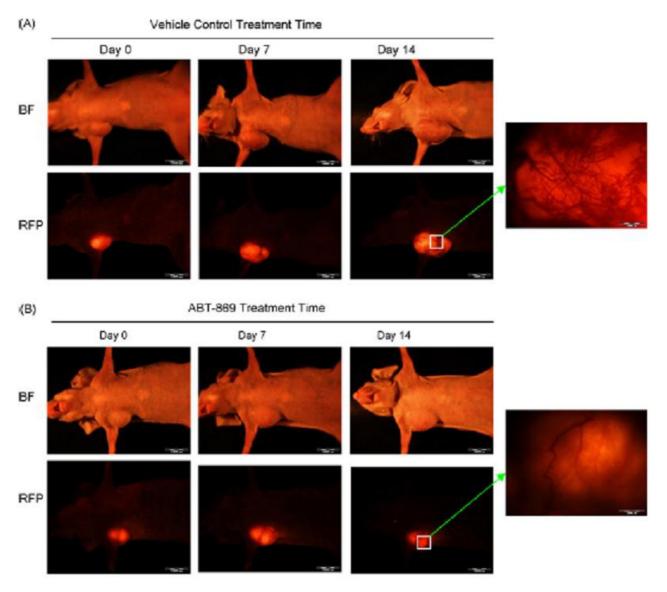


Figure 7
Sequential real-time whole-body fluorescence imaging of HL60-RFP tumor growth in living mice. (A) Mice were treated with vehicle control. (B) Mice treated with ABT-869 (15 mg/kg/day). Arrow-pointed pictures show the direct view of distribution of blood vessel network on the tumor surface in the two representative mice. There is less of a tumor vessel network in ABT-869 treated mice. BF: bright field channel. RFP: RFP channel (The picture is modified from Leukemia Research 2008; 32:1091–1100 with permission) [32].

some of the mechanisms that could potentially be used by leukemia cells to overcome the cytotoxic effect under long-term use of ABT-869. Three resistant cell lines (designated as MV4–11-R1, -R2, -R3) were developed by over three-month co-culture of the human leukemia cell line, MV4–11 (AML, both alleles FLT3-ITD) with increasing concentrations of ABT-869 [50]. These resistant lines are much less sensitive to ABT-869-medidated cell proliferation inhibition and apoptosis, but also are cross-resistant to structurally unrelated FLT3 inhibitors (AG1296,

SU5416 and FLT3 inhibitor III). No point mutation is found in the FLT3 kinase domain in all 3 resistant lines [50]. Low density array analysis reveals that a total of 61 genes are differentially expressed more than 2-fold between the 3 resistant and parental MV4–11 cells. Interestingly, MV4–11-R cells over-express FLT3 ligand (FLT3LG) and BIRC5 (Survivin), while down-regulate the suppressor of cytokine signaling (SOCS) family (SOCS-1, -2, -3) [50]. The C-terminal domain of SOCS proteins acts as an adapter targeting kinase receptor complex for ubiq-

uitination and subsequent proteasome-mediated degradation [51]. The SOCS family also is an important negative regulator of STAT pathways [51,52]. In MV4–11-R cells, hypermethylation silencing of SOCS genes leads to reactivation of STAT pathway activities, as evidenced by increasing levels of phosphorylation of STAT1 protein (p-STAT1), p-STAT3 and p-STAT5 [50].

Membrane-bound and soluble forms of FLT3 ligand are both biologically active [53]. FLT3 ligand plays an important role in survival, proliferation, and differentiation of hematopoietic stem and progenitor cells (HSPC) [54,55]. It has been demonstrated that the autocrine FLT3LG/FLT3 loop promotes proliferation and prevents apoptosis of primary AML blasts and AML cell lines.[56,57] Stimulation of MV4-11 cells with extra FLT3 ligand either by directly adding to the culture medium or by using conditioned medium harvested from MV4-11-R cells can further increase p-STAT1, p-STAT3, p-STAT5, as well as the expression of survivin [50], which correlate with resistance to ABT-869 and other FLT3 inhibitors (AG1296, SU5416 and FLT3 inhibitor III). On the contrary, blocking FLT3 ligand with a FLT3 ligand neutralizing antibody enhances ABT-869-induced apoptosis in MV4-11-R cells [50]. Collectively, these results indicate a prominent role of FLT3 ligand in mediating the resistance to FLT3 inhibitors.

Survivin (encoded by BIRC5), the smallest member of the inhibitor of apoptosis protein (IAP) family, has been regarded as one of the classic fetal oncoproteins [58-61]. Survivin stabilizes X-linked IAP (XIAP), another member of IAP family, against proteasomal degradation to protect cells from apoptosis [62]. To demonstrate the critical role of survivin in the regulation of resistance in MV4–11-R cells, a pool of shRNA was used to specially target survivin. Silencing survivin remarkably potentiates ABT-869-induced apoptosis in MV4–11-R cells when compared to control shRNA treatment. In contrast, forced expression of survivin in MV4–11 cells leads to resistant to ABT-869 and other FLT3 inhibitors [50].

After screening for compounds which could potentially reverse the resistance phenotype in MV4–11R, Indirubin derivative (IDR) E804 was identified. As an inhibitor of the SRC-STAT3 pathway [63], IDR E804 shows potent efficacy in re-sensitizing MV4–11-R to ABT-869. IDR E804 treatment dose-dependently induces MV4–11-R cells to undergo apoptosis and inhibits the expression of p-STAT1, p-STAT3, p-STAT5 as well as completely abolishes survivin expression [50]. In the presence of a sub-toxic concentration (2 nM) of IDR E804, the IC50 value of ABT-869 in MV4–11-R decreased from 52 to 6 nM. The combination of ABT-869 and IDR E804 also achieves better anti-

tumor effect than either single agent treatment in a MV4–11-R mouse xenograft model [50].

In summary, over expression of FLT3 ligand, methylation silencing of the SOCS family and overexpression of survivin all together integrate leading aberrant STAT signaling activity and contribute to resistance to FLT3 inhibitors. The discovery of this novel mechanism of resistance to FLT3 inhibitors, as described in Figure 8, could help develop new anti-leukemic agents or uncover compelling combinations. Combination of FLT3 inhibitors with compounds targeting the STAT pathway or survivin may represent a therapeutic strategy to minimize resistance or resensitize resistant cells to FLT3 inhibitors in AML patients with FLT3-ITD mutation.

First in Man (FIM) and phase I study

In 2006, Abbott made a strategic decision and partnered with the clinical team at National University Hospital in Singapore and conducted the first in man study for ABT-869. The first in man study was started in patients with solid malignancies refractory to or for which no standard effective therapy exists who were enrolled in escalating dose cohorts and treated with oral ABT-869 once daily continuously. This study was designed as a single-arm, open-label Phase I trial and was conducted in three segments in order to determine the maximum tolerable dose (MTD), tolerability, and pharmacodynamics of a lower dose cohort to better define dose-effect relationships. ABT-869 lacks high aqueous solubility, therefore, the study drug was diluted in 60 mLs of Ensure Plus®. Preliminary PK at doses of 10 mg showed a modest correlation

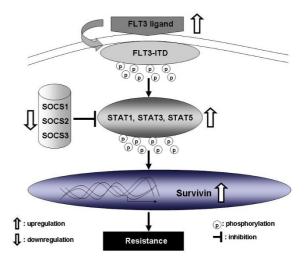


Figure 8
A model of enhanced STAT activation and overexpression of survivin leading to resistant phenotype in MV4–II-R cells. (Modified with permission from Blood journal) [50].

between oral clearance and body-weight; thus subsequent dose escalations in segment A were based on bodyweight. The most common drug-related adverse events were fatigue, proteinuria, hypertension, myalgia, skin toxicity (hand and foot blisters) and oral hypersensitivity, and these toxicities increased in frequency and intensity with increasing doses. The maximal tolerated dose (MTD) was determined to be 0.3 mg/kg/day. In general, the treatments are well tolerated in this patient population with either refractory disease or no standard therapy.

The treatment response of this phase I trial is encouraging. Three (10%) out of 29 patients achieved partial response (PR); two had non-small cell lung cancer (NSCLC) treated at 0.3 mg/kg/day and 10 mg/day respectively, and one had colorectal cancer (CRC) treated at 0.1 mg/kg/day. An additional sixteen patients had stable disease lasting longer than 12 weeks, among which were patients with CRC (5), NSCLC (2), ovarian cancer (2), hepatocellular carcinoma (HCC) (2) and neuroendocrine tumour (2).

Tumor cavitation in the lungs and reduction of contrast enhancement in tumor on post-treatment CT scans after ABT-869 treatment suggesting central necrosis supported antiangiogenic activity, and has been observed with other VEGF antagonists (Figure 9). Prolonged stable disease lasting more than 12 months with minimal toxicity was observed in four patients; alveolar soft part sarcoma (27 months), CRC (19 months), HCC (17 months), and renal

cell carcinoma (18 months) [64]. The response to ABT-869 observed in multiple tumor types suggests that histological different types of cancer could share the same dysregulated signaling pathway(s) and the rationale of multitargeted approach may be necessary for solid tumors.

Extensive pharmacodynamic analyses were performed with this phase I trial. Exposures of ABT-869 (AUC from 0-24 h) from this trial were similar between Asian and Caucasian populations (2.7 vs. 2.3 μg·h/mL, respectively) and met the exposure targets derived from nonclinical efficacy studies [18,64]. Dynamic contrast enhanced-MRI (DCE-MRI) showed dose-dependent reduced tumor vascular permeability that correlated with drug exposure. Circulating endothelial cells (CECs) were significantly reduced $(9.6 \pm 7.0/\mu L \text{ vs. } 16.5 \pm 13.4/\mu L, p = 0.007)$ and vascular endothelial factor was increased (126.3 ± 104.4 pg/mL vs. 74.2 ± 82.2 pg/mL, p = 0.004) by day 15 of treatment (0.25 mg/kg) [64]. The biomarker evidence of antiangiogenic activity and DCE-MRI evidence of tumor antiangogenesis are consistent with proof of target inhibition and can be translated to observed promising clinical activity.

A multi-center phase I study was also initiated in patients with refractory or relapsed AML or myelodysplastic syndrome (MDS) as FLT-3 is an obvious therapeutic target of ABT-869. Based on our pre-clinical study [22], the trial was designed as two stages with initial monotherapy and

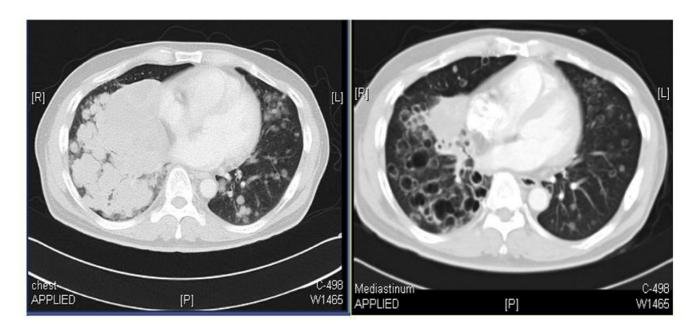


Figure 9
Computed tomography scan of tumor response and cavitation of lesions in a patient with metastatic lung carcinoma showing cavitation after 2 treatment periods. (with permission from Journal of Clinical Oncology) [64].

later in combination with Ara-C. Specifically, based on our pre-clinical combination sequence data, ABT-869 will be given after the completion of Ara-C at each cycle.

Current ongoing clinical trials

The promising anti-cancer properties of ABT-869 identified at the early phase trial facilitate further clinical development of this novel agent. In June 2007, Abbott and Genentech Inc. formed collaboration for the global research, development and commercialization for ABT-869. Phase II clinical trials evaluating ABT-869 for advanced or metastatic hepatocellular carcinoma, metastatic breast cancer, metastatic colorectal cancer, metastatic non-small cell lung cancer, and advanced renal cell carcinoma are ongoing. A summary of current ABT-869 clinical trials listed on the National Institutes of Health Website is shown in Table 2.

Preliminary clinical data on single agent ABT-869 was presented in the 2009 ASCO annual meeting. Encouraging clinical activity has been observed in non-small cell lung cancer (NSCLC) and advanced hepatocellular carcinoma

(HCC) trials as well as in a renal cell carcinoma (RCC) trial after Sunitinib failure [65-67]. However, additional studies are required to determine the optimal dosing strategy especially in RCC and HCC patient population as frequent dose interruption or reduction was observed. In the NSCLC trial, two different doses were tested (0.10 mg/kg and 0.25 mg/kg), and preliminary data did not show significant difference in OS and PFS between these two arms. Furthermore, current pharmacokinetic analysis indicates that body weight does not significantly impact exposure suggesting that a fixed dosing strategy may be appropriate [68].

Conclusions and future directions

In summary, ABT-869 is a novel inhibitor that simultaneously provides potent and selective inhibition of the VEGFR and PDGFR kinase families and has demonstrated activity in patients with solid tumors who failed standard regimen. Optimal dosing and scheduling are being investigated and the potent *in vivo* angiogenesis effect has already produced a promising clinical response in early phase clinical development.

Table 2: Current listed clinical trials on ABT-869

Trial title	Enrollment	Trial design	Last verified	Recruitment	Start date
Phase 2 Study of ABT-869 in Combination With Paclitaxel Versus Paclitaxel Alone as First Line Treatment For Metastatic Breast Cancer	102	RDBT, MC	April 2009	Recruiting	March 2008
Phase 2 Study of ABT-869 in Advanced Hepatocellular Carcinoma (HCC)	44	RDBT, MC	March 2009	Active, not recruiting	August 2007
Study of ABT-869 in Combination With Tarceva in Subjects With Solid Tumors	0	W	January 2009	Withdrawn	September 2008
Phase I Study of ABT-869 in Subjects With Solid Tumors	24	Conducted in Japan	March 2009	Recruiting	September 2008
Phase 2 Study of ABT-869 in Subjects With Advanced Non-Small Cell Lung Cancer (NSCLC)	139	RUO, MC	March 2009	Active, not recruiting	August 2007
Phase 2 Study of ABT-869 in Combination With mFOLFOX6 Versus Bevacizumab in Combination With mFOLFOX6 as Second Line Treatment for Advanced Colorectal Cancer	102	RUO, MC	April 2009	Recruiting	August 2008
Phase 2 Study of Carboplatin/Paclitaxel in Combination With ABT-869 in Subjects With Advanced or Metastatic Non-Small Cell Lung Cancer (NSCLC)	80	RDBT, MC	April 2009	Recruiting	June 2008
Phase 2 Study of ABT-869 in Subjects With Advanced Renal Cell Carcinoma Who Have Previously Received Treatment With Sunitinib	53	Open label, NR	April 2009	Active, not recruiting	August 2007
Phase 2 Study of Oxaliplatin, Fluorouracil, Leucovorin and ABT-869 or Bevacizumab as Second-Line Therapy in Treating Patients With Locally Recurrent or Metastatic Colorectal Cancer	0	Single center	October 2008	Not yet recruiting	October 2008
Phase I Pharmacokinetic Study To Evaluate Effect of Food and Diurnal Variation on ABT-869	12	Single center	March 2009	Recruiting	February 2009

Data compiled from http://www.clinicaltrials.org

RDBT: Randomized, placebo-controlled, double blind trial

MC: Multicenter

W: Withdrawn prior to recruitment

RUO: Randomized, uncontrolled, open label

NR: Non-Randomized

Based on the Population PK analysis presented in an abstract [68], ABT-869 PK fits one-compartment model with first order absorption and elimination. Race, sex and impaired renal function do not appear to significantly affect PK. In addition, body weight does not significantly impact exposure suggesting that a fixed dosing strategy may be appropriate.

The reported side effects such as fatigue, proteinuria, hypertension, myalgia, skin toxicity (hand and foot blisters) are similar to commonly described toxicity in other FDA approved oral tyrosine kinase inhibitors such as Sunitinib. Long term dosing of ABT-869 did not appear to pose problems of cumulative toxicity in patients who received more than a year of dosing. The nonclinical studies on combination therapies have demonstrated synergy and are likely to be more effective than monotherapy. Clinical studies of ABT-869 in combination with chemotherapy or other novel targeted therapies, will further our understanding of how to optimize this exciting new therapy. The recent identification of the critical role of survivin in the regulation of ABT-869 resistance is interesting and is therapeutically relevant. Mechanisms of resistance to ABT-869 remain under active investigation.

Competing interests

DHA is an employee of Abbott laboratories. The other authors declared no conflict of interest.

Authors' contributions

JZ wrote the review of pre-clinical studies. BCG contributed the phase I trial data, and DHA contributed non-clinical studies, discussion and revision. CSC conceived of the study, wrote the review of clinical trials and organized the manuscript. All authors read and approved the final manuscript.

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References

- Blume-Jensen P, Hunter T: Oncogenic kinase signalling. Nature 2001, 411:355-65.
- den Hertog J, Ostman A, Bohmer FD: Protein tyrosine phosphatases: regulatory mechanisms. Febs J 2008, 275:831-47.
- Rosnet O, Birnbaum D: Hematopoietic receptors of class III receptor-type tyrosine kinases. Crit Rev Oncog 1993, 4:595-613.
- Small D, Levenstein M, Kim E, Carow C, Amin S, Rockwell P, Witte L, Burrow C, Ratajczak MZ, Gewirtz AM, et al.: STK-I, the human homolog of Flk-2/Flt-3, is selectively expressed in CD34+ human bone marrow cells and is involved in the proliferation of early progenitor/stem cells. Proc Natl Acad Sci USA 1994, 91:459-63.
- Lyman SD, James L, Johnson L, Brasel K, de Vries P, Escobar SS, Downey H, Splett RR, Beckmann MP, McKenna HJ: Cloning of the

- human homologue of the murine flt3 ligand: a growth factor for early hematopoietic progenitor cells. Blood 1994, 83:2795-801
- Nakao M, Yokota S, Iwai T, Kaneko H, Horiike S, Kashima K, Sonoda Y, Fujimoto T, Misawa S: Internal tandem duplication of the flt3 gene found in acute myeloid leukemia. Leukemia 1996, 10:1911-8.
- Gilliland DG, Griffin JD: The roles of FLT3 in hematopoiesis and leukemia. Blood 2002, 100:1532-42.
- Stirewalt DL, Radich JP: The role of FLT3 in haematopoietic malignancies. Nat Rev Cancer 2003, 3:650-65.
- Levis M, Small D: FLT3: IT Does matter in leukemia. Leukemia 2003. 17:1738-52.
- Sternberg DW, Licht JD: Therapeutic intervention in leukemias that express the activated fms-like tyrosine kinase 3 (FLT3): opportunities and challenges. Curr Opin Hematol 2005, 12:7-13.
- Ferrara N, Gerber HP, LeCouter J: The biology of VEGF and its receptors. Nat Med 2003, 9:669-76.
- 12. Bergers G, Benjamin LE: Tumorigenesis and the angiogenic switch. Nat Rev Cancer 2003, 3:401-10.
- Zhou J, Mauerer K, Farina L, Gribben JG: The role of the tumor microenvironment in hematological malignancies and implication for therapy. Front Biosci 2005, 10:1581-96.
- Dias S, Hattori K, Zhu Z, Heissig B, Choy M, Lane W, Wu Y, Chadburn A, Hyjek E, Gill M, Hicklin DJ, Witte L, Moore MA, Rafii S: Autocrine stimulation of VEGFR-2 activates human leukemic cell growth and migration. J Clin Invest 2000, 106:511-21.
- Fiedler W, Graeven U, Ergun S, Verago S, Kilic N, Stockschlader M, Hossfeld DK: Vascular endothelial growth factor, a possible paracrine growth factor in human acute myeloid leukemia. Blood 1997, 89:1870-5.
- List AF, Glinsmann-Gibson B, Stadheim C, Meuillet EJ, Bellamy W, Powis G: Vascular endothelial growth factor receptor-I and receptor-2 initiate a phosphatidylinositide 3-kinase-dependent clonogenic response in acute myeloid leukemia cells. Exp Hematol 2004, 32:526-35.
- Santos SC, Días S: Internal and external autocrine VEGF/KDR loops regulate survival of subsets of acute leukemia through distinct signaling pathways. Blood 2004, 103:3883-9.
- Albert DH, Tapang P, Magoc TJ, Pease LJ, Reuter DR, Wei RQ, Li J, Guo J, Bousquet PF, Ghoreishi-Haack NS, Wang B, Bukofzer GT, Wang YC, Stavropoulos JA, Hartandi K, Niquette AL, Soni N, Johnson EF, McCall JO, Bouska JJ, Luo Y, Donawho CK, Dai Y, Marcotte PA, Glaser KB, Michaelides MR, Davidsen SK: Preclinical activity of ABT-869, a multitargeted receptor tyrosine kinase inhibitor. Mol Cancer Ther 2006, 5:995-1006.
- 19. Dai Y, Hartandi K, Ji Z, Ahmed AA, Albert DH, Bauch JL, Bouska JJ, Bousquet PF, Cunha GA, Glaser KB, Harris CM, Hickman D, Guo J, Li J, Marcotte PA, Marsh KC, Moskey MD, Martin RL, Olson AM, Osterling DJ, Pease LJ, Soni NB, Stewart KD, Stoll VS, Tapang P, Reuter DR, Davidsen SK, Michaelides MR: Discovery of N-(4-(3-amino-1H-indazol-4-yl)phenyl)-N'-(2-fluoro-5-methylphenyl)urea (ABT-869), a 3-aminoindazole-based orally active multitargeted receptor tyrosine kinase inhibitor. J Med Chem 2007, 50:1584-97.
- Guo J, Marcotte PA, McCall JO, Dai Y, Pease LJ, Michaelides MR, Davidsen SK, Glaser KB: Inhibition of phosphorylation of the colony-stimulating factor-I receptor (c-Fms) tyrosine kinase in transfected cells by ABT-869 and other tyrosine kinase inhibitors. Mol Cancer Ther 2006, 5:1007-13.
- Li TWJP: Cellular activity of ABT-869 against colony-stimulating-factor-1 receptor (CSF-1R) in macrophage-like cell lines [abstract]. In Proceedings of the 100th Annual Meeting of the American Association for Cancer Research Denver, CO: Philadelphia (PA): AACR; 2009. Abstract nr 1789
- Zhou J, Pan M, Xie Z, Loh SL, Bi C, Tai YC, Lilly M, Lim YP, Han JH, Glaser KB, Albert DH, Davidsen SK, Chen CS: Synergistic antileukemic effects between ABT-869 and chemotherapy involve downregulation of cell cycle-regulated genes and c-Mos-mediated MAPK pathway. Leukemia 2008, 22:138-46.
- Donawho C HJ, Wang YC, Bukofzer G, Dai Y, Davidsen S, Wang B, Schlessinger S, Frost D: The RTK inhibitor ABT-869, alone and in combination with paclitaxel and/or zoledronic acid, demonstrates significant reduction in the development of both osteoblastic (LuCap 23.1) and osteolytic (PC3-M-Luciferase) tumors intratibially. AACR Meeting Abstracts 2007 2007:C204.

- Jasinghe VJ, Xie Z, Zhou J, Khng J, Poon LF, Senthilnathan P, Glaser KB, Albert DH, Davidsen SK, Chen CS: ABT-869, a multi-targeted tyrosine kinase inhibitor, in combination with rapamycin is effective for subcutaneous hepatocellular carcinoma xenograft. J Hepatol 2008, 49:985-97.
- Li L, Lin X, Shoemaker AR, Albert DH, Fesik SW, Shen Y: Hypoxiainducible factor-I inhibition in combination with temozolomide treatment exhibits robust antitumor efficacy in vivo. Clin Cancer Res 2006, 12:4747-54.
- Kim KT, Baird K, Ahn JY, Meltzer P, Lilly M, Levis M, Small D: Pim-I is up-regulated by constitutively activated FLT3 and plays a role in FLT3-mediated cell survival. Blood 2005, 105:1759-67.
- Shankar DB, Li J, Tapang P, Owen McCall J, Pease LJ, Dai Y, Wei RQ, Albert DH, Bouska JJ, Osterling DJ, Guo J, Marcotte PA, Johnson EF, Soni N, Hartandi K, Michaelides MR, Davidsen SK, Priceman SJ, Chang JC, Rhodes K, Shah N, Moore TB, Sakamoto KM, Glaser KB: ABT-869, a multitargeted receptor tyrosine kinase inhibitor: inhibition of FLT3 phosphorylation and signaling in acute myeloid leukemia. Blood 2007, 109:3400-8.
- 28. Brown P, Levis M, McIntyre E, Griesemer M, Small D: Combinations of the FLT3 inhibitor CEP-701 and chemotherapy synergistically kill infant and childhood MLL-rearranged ALL cells in a sequence-dependent manner. Leukemia 2006, 20:1368-76.
- Fiedler W, Mesters R, Tinnefeld H, Loges S, Staib P, Duhrsen U, Flasshove M, Ottmann OG, Jung W, Cavalli F, Kuse R, Thomalla J, Serve H, O'Farrell AM, Jacobs M, Brega NM, Scigalla P, Hossfeld DK, Berdel WE: A phase 2 clinical study of SU5416 in patients with refractory acute myeloid leukemia. Blood 2003, 102:2763-7.
- O'Farrell AM, Abrams TJ, Yuen HA, Ngai TJ, Louie SG, Yee KW, Wong LM, Hong W, Lee LB, Town A, Smolich BD, Manning WC, Murray LJ, Heinrich MC, Cherrington JM: SU11248 is a novel FLT3 tyrosine kinase inhibitor with potent activity in vitro and in vivo. Blood 2003, 101:3597-605.
- Roboz GJ, Giles FJ, List AF, Cortes JE, Carlin R, Kowalski M, Bilic S, Masson E, Rosamilia M, Schuster MW, Laurent D, Feldman EJ: Phase I study of PTK787/ZK 22 a small molecule tyrosine kinase receptor inhibitor, for the treatment of acute myeloid leukemia and myelodysplastic syndrome. Leukemia 2584, 20:952-7.
- Zhou J, Khng J, Jasinghe VJ, Bi C, Neo CH, Pan M, Poon LF, Xie Z, Yu H, Yeoh AE, Lu Y, Glaser KB, Albert DH, Davidsen SK, Chen CS: In vivo activity of ABT-869, a multi-target kinase inhibitor, against acute myeloid leukemia with wild-type FLT3 receptor. Leuk Res 2008, 32:1091-100.
- Semenza GL: Angiogenesis in ischemic and neoplastic disorders. Annu Rev Med 2003, 54:17-28.
- Yamaguchi R, Yano H, lemura A, Ogasawara S, Haramaki M, Kojiro M: Expression of vascular endothelial growth factor in human hepatocellular carcinoma. Hepatology 1998. 28:68-77
- hepatocellular carcinoma. Hepatology 1998, 28:68-77.
 35. Baldo P, Cecco S, Giacomin E, Lazzarini R, Ros B, Marastoni S: mTOR pathway and mTOR inhibitors as agents for cancer therapy. Curr Cancer Drug Targets 2008, 8:647-65.
- Schmelzle T, Hall MN: TOR, a central controller of cell growth. Cell 2000, 103:253-62.
- Yap TA, Garrett MD, Walton MI, Raynaud F, de Bono JS, Workman P: Targeting the PI3K-AKT-mTOR pathway: progress, pitfalls, and promises. Curr Opin Pharmacol 2008, 8:393-412.
- Llovet JM, Bruix J: Molecular targeted therapies in hepatocellular carcinoma. Hepatology 2008, 48:1312-27.
- Tanaka S, Arii S: Molecularly targeted therapy for hepatocellular carcinoma. Cancer Sci 2009, 100:1-8.
- Guba M, von Breitenbuch P, Steinbauer M, Koehl G, Flegel S, Hornung M, Bruns CJ, Zuelke C, Farkas S, Anthuber M, Jauch KW, Geissler EK: Rapamycin inhibits primary and metastatic tumor growth by antiangiogenesis: involvement of vascular endothelial growth factor. Nat Med 2002, 8:128-35.
- Sahin F, Kannangai R, Adegbola O, Wang J, Su G, Torbenson M: mTOR and P70 S6 kinase expression in primary liver neoplasms. Clin Cancer Res 2004, 10:8421-5.
- DeAngelo DJ, Stone RM, Heaney ML, Nimer SD, Paquette RL, Klisovic RB, Caligiuri MA, Cooper MR, Lecerf JM, Karol MD, Sheng S, Holford N, Curtin PT, Druker BJ, Heinrich MC: Phase I clinical results with tandutinib (MLN518), a novel FLT3 antagonist, in patients with acute myelogenous leukemia or high-risk myelodysplastic syndrome: safety, pharmacokinetics, and pharmacodynamics. Blood 2006, 108:3674-81.

- 43. Fiedler W, Serve H, Dohner H, Schwittay M, Ottmann OG, O'Farrell AM, Bello CL, Allred R, Manning WC, Cherrington JM, Louie SG, Hong W, Brega NM, Massimini G, Scigalla P, Berdel WE, Hossfeld DK: A phase I study of SUI1248 in the treatment of patients with refractory or resistant acute myeloid leukemia (AML) or not amenable to conventional therapy for the disease. Blood 2005, 105:986-93.
- Knapper S, Burnett AK, Littlewood T, Kell WJ, Agrawal S, Chopra R, Clark R, Levis MJ, Small D: A phase 2 trial of the FLT3 inhibitor lestaurtinib (CEP701) as first-line treatment for older patients with acute myeloid leukemia not considered fit for intensive chemotherapy. Blood 2006, 108:3262-70.
- Small D: FLT3 mutations: biology and treatment. Hematology Am Soc Hematol Educ Program 2006:178-84.
- Druker BJ, Sawyers CL, Čapdeville R, Ford JM, Baccarani M, Goldman JM: Chronic myelogenous leukemia. Hematology (Am Soc Hematol Educ Program) 2001:87-112.
- Gorre ME, Mohammed M, Ellwood K, Hsu N, Paquette R, Rao PN, Sawyers CL: Clinical resistance to STI-571 cancer therapy caused by BCR-ABL gene mutation or amplification. Science 2001, 293:876-80.
- Heidel F MF, Breitenbucher F, Dove S, Huber C, Bohmer FD, Ficher T: Bis(1H-indol-2-yl)methanones are effective inhibitors of mutated FLT3 tyrosine kinase, overcome resistance to PKC421A in vitro and show synergy with chemotherapy. Blood 2006, 108:.
- Piloto O, Wright M, Brown P, Kim KT, Levis M, Small D: Frolonged exposure to FLT3 inhibitors leads to resistance via activation of parallel signaling pathways. Blood 2007, 109:1643-52.
- Zhou J, Bi C, Janakakumara JV, Liu SC, Chng WJ, Tay KG, Poon LF, Xie Z, Palaniyandi S, Yu H, Glaser KB, Albert DH, Davidsen SK, Chen CS: Enhanced activation of STAT pathways and overexpression of survivin confer resistance to FLT3 inhibitors and could be therapeutic targets in AML. Blood 2009, 113:4052-62.
 Wormald S, Hilton DJ: The negative regulatory roles of sup-
- Wormald S, Hilton DJ: The negative regulatory roles of suppressor of cytokine signaling proteins in myeloid signaling pathways. Curr Opin Hematol 2007, 14:9-15.
- Bromberg J: Stat proteins and oncogenesis. J Clin Invest 2002, 109:1139-42.
- Hannum C, Culpepper J, Campbell D, McClanahan T, Zurawski S, Bazan JF, Kastelein R, Hudak S, Wagner J, Mattson J, et al.: Ligand for FLT3/FLK2 receptor tyrosine kinase regulates growth of haematopoietic stem cells and is encoded by variant RNAs. Nature 1994, 368:643-8.
- Rusten LS, Lyman SD, Veiby OP, Jacobsen SE: The FLT3 ligand is a direct and potent stimulator of the growth of primitive and committed human CD34+ bone marrow progenitor cells in vitro. Blood 1996, 87:1317-25.
- Dehmel U, Zaborski M, Meierhoff G, Rosnet O, Birnbaum D, Ludwig WD, Quentmeier H, Drexler HG: Effects of FLT3 ligand on human leukemia cells. I. Proliferative response of myeloid leukemia cells. Leukemia 1996, 10:261-70.
- Zheng R, Levis M, Piloto O, Brown P, Baldwin BR, Gorin NC, Beran M, Zhu Z, Ludwig D, Hicklin D, Witte L, Li Y, Small D: FLT3 ligand causes autocrine signaling in acute myeloid leukemia cells. Blood 2004, 103:267-74.
- 57. Scheijen B, Ngo HT, Kang H, Griffin JD: FLT3 receptors with internal tandem duplications promote cell viability and proliferation by signaling through Foxo proteins. *Oncogene* 2004, 23:3338-49.
- Ambrosini G, Adida C, Altieri DC: A novel anti-apoptosis gene, survivin, expressed in cancer and lymphoma. Nat Med 1997, 3:917-21.
- Zaffaroni N, Pennati M, Daidone MG: Survivin as a target for new anticancer interventions. J Cell Mol Med 2005, 9:360-72.
- Fukuda S, Pelus LM: Survivin, a cancer target with an emerging role in normal adult tissues. Mol Cancer Ther 2006, 5:1087-98.
- 61. Altieri DC: Survivin, cancer networks and pathway-directed drug discovery. Nat Rev Cancer 2008, 8:61-70.
- Altieri DC: Validating survivin as a cancer therapeutic target. Nat Rev Cancer 2003, 3:46-54.
- Nam S, Buettner R, Turkson J, Kim D, Cheng JQ, Muehlbeyer S, Hippe F, Vatter S, Merz KH, Eisenbrand G, Jove R: Indirubin derivatives inhibit Stat3 signaling and induce apoptosis in human cancer cells. Proc Natl Acad Sci USA 2005, 102:5998-6003.

- 64. Wong Cl, Koo TS, Soo R, Hartono S, Thng CH, McKeegan E, Yong WP, Chen CS, Lee SC, Wong J, Lim R, Sukri N, Lim SE, Ong AB, Steinberg J, Gupta N, Pradhan R, Humerickhouse R, Goh BC: Phase I and biomarker study of ABT-869, a multiple receptor tyrosine kinase inhibitor, in patients with refractory solid malignancies. Journal of Clinical Oncology 2009 in press.
- cies. Journal of Clinical Oncology 2009 in press.

 65. Toh H, Chen P, Carr BI, Knox JJ, Gill S, Steinberg J, Carlson DM, Qian J, Qin Q, Yong W: A phase II study of ABT-869 in hepatocellular carcinoma (HCC): Interim analysis. J Clin Oncol 2009, 27(20e15-e17 [http://meeting.ascopubs.org/cgi/content/abstract/27/15S/4581].
- 66. Tannir N, Wong Y, Kollmannsberger C, Ernstoff MS, Perry DJ, Appleman LJ, Posadas E, Qian J, Ricker JL, Michaelson DM: Phase II trial of ABT-869 in advanced renal cell cancer (RCC) after sunitinib failure: Efficacy and safety results. J Clin Oncol 2009, 27:15s [http://meeting.ascopubs.org/cgi/content/abstract/27/15S/5036].
- 67. Tan E, Salgia R, Besse B, Goss G, Gandara DR, Hanna N, Steinberg J, Steinberg J, Qian J, Carlson DM, Soo R: **ABT-869** in non-small cell lung cancer (NSCLC): Interim results. Clin Oncol 2009, 27:15s [http://meeting.ascopubs.org/cgi/content/abstract/27/15S/8074].
- [http://meeting.ascopubs.org/cgi/content/abstract/27/15S/8074].
 68. Gupta N, Diderichsen PM, Steinberg J, Ricker JL, Humerickhouse R, Awni W, Pradhan R: Population pharmacokinetic (PK) analysis of ABT-869 in solid tumors and acute myelogenous leukemia (AML) patients. J Clin Oncol 2009, 27:15s [http://meeting.ascopubs.org/cgi/content/abstract/27/15S/3567].

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